




**Expression of Uncertainty in Qualitative Analysis**

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**Introduction**

- What is “Qualitative Analysis”?
- Characterising uncertainty and method performance
- Qualitative response dependent on a concentration
- What can we expect from labs?



## What is qualitative analysis?



- “Classification according to specific criteria”
  - “Above” or “Below” a limit
  - “Within Spec.”
  - “Red”
  - Classification into ranges (<2; 2-5; 5-10; >10)
  - Molecular species by NMR, IR, MS.....
  - Material or ingredient (“Rubber”, “Fat”...)
  - Origin or authenticity



## Expression of uncertainty in qualitative analysis



- False response rates
  - What is a false response rate?
  - How is it determined?
- Alternative expressions of method performance or uncertainty
- Logistic regression for modelling performance

**NOTE**  
Current literature refers to  
“nominal properties”



## False response rates and derived indicators



		Actual (True) value	
		Negative	Positive
Observed	Negative	TN	FN
	Positive	FP	TP

## Alternative performance indicators (Single laboratory)



Reliability Measure	Formula
False positive rate	$FP / (TN + FP)$
False negative rate	$FN / (TP + FN)$
Sensitivity	$TP / (TP + FN)$
Specificity	$TN / (TN + FP)$
Positive predictive value	$TP / (TP + FP)$
Efficiency	$(TP + TN) / (TP + TN + FP + FN)$
Youden Index	Sensitivity + Specificity - 100
Likelihood ratio	$(1 - \text{False neg. rate}) / (\text{False pos. rate})$
Bayes posterior probability	Bayes rule (requires 'prior') - valuable for cumulative data

Proportion of positives that are correct

Uncertainty about the result



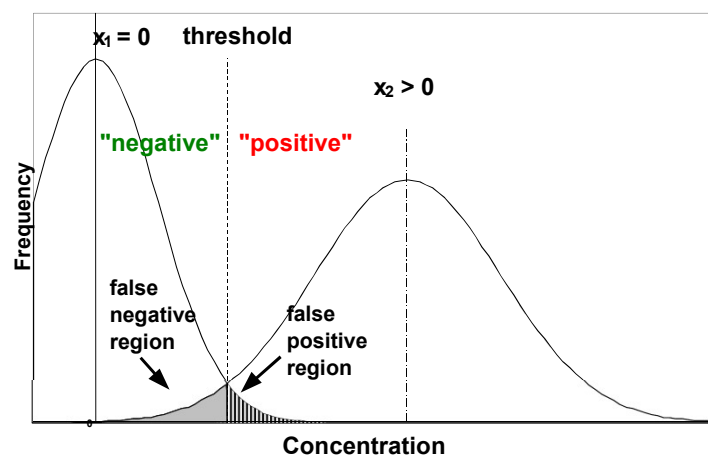
## False response rates - how much data?



- Observed: 7/126 (5.6%)
- 95% confidence interval (binomial)
  - 1.6% to 9.5%
- 95% CI proportional to  $1/\sqrt{n_{\text{obs}}}$ 
  - needs a LOT of false responses for precise figures
  - but false responses are rare for good methods....
- Most useful direct studies are 'worst case' or near 50% false response levels



## False responses: Estimation from thresholds



## False responses: From probabilities



- Spectroscopic identification study
  - S.L.R. Ellison, S.L. Gregory, *Anal. Chim. Acta.*, 1998 **370** 181.
- Calculated chance FT-IR match probabilities
  - probabilities based on “match-binning” - hits within set distance
  - required hypergeometric distribution ( $n$  matches of  $m$  taken from population)
- Compared with actual hits on IR database



## False responses: From probabilities



- Theoretical predictions very sensitive to probability assumptions
  - 10% changes in  $p$  make large differences in predictions
- Best performance within factor of 3-10
  - (Improved over binomial probabilities by  $>10^6$ )
- Probability information must be excellent for good predictions



## False response rates from databases



- Most spectral databases contain 1 of each material
  - most populations do not!
- Population data must account for sub-populations
  - cf. DNA profiles for racially inhomogeneous populations



## Using Logistic Regression



- Logistic regression models *probability* as a function of a continuous variable

$$E(Y | x) = \frac{e^{\beta_0 + \beta_1 x}}{1 + e^{\beta_0 + \beta_1 x}}$$

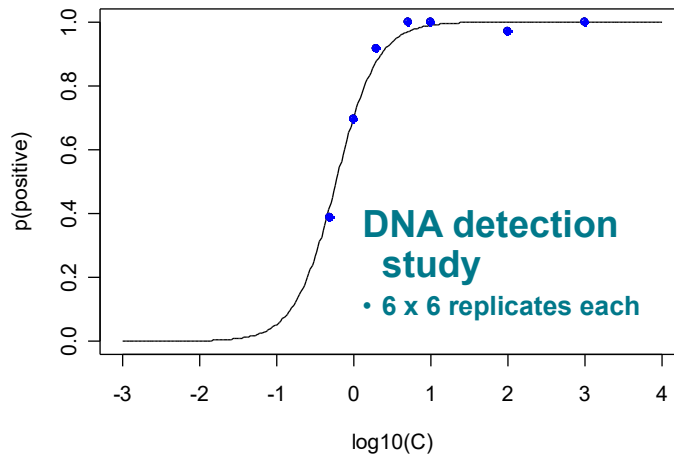
- Example:
  - p(DNA found) vs DNA concentration



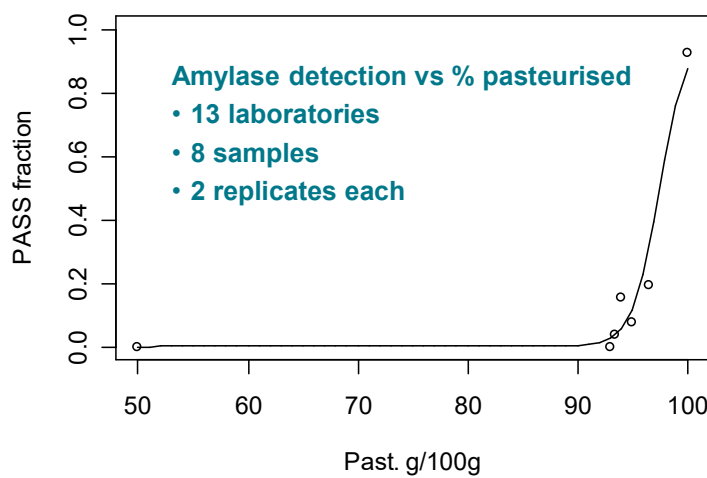
## Logistic regression and performance assessment



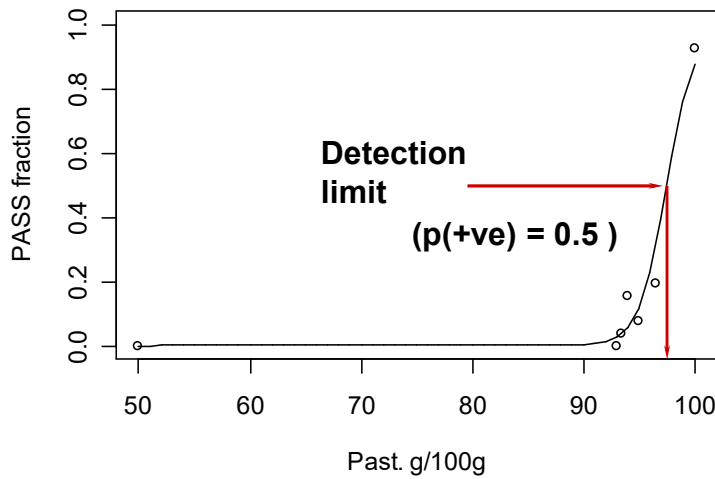
Logistic Regression: p(positive) by log<sub>10</sub>(C)



## Logistic regression and method performance



## Logistic regression and method performance



## Problems for qualitative “uncertainty”



- Hard to estimate low false response rates
  - May take hundreds of experiments
- Harder to estimate population probabilities
- Harder still to evaluate joint probabilities
  - ... and these have large effects on calculation
- Prior probabilities are very rarely available





## Recommendations\*



- It is realistic to expect that testing laboratories have qualitative test method parameters (conditions of testing) under adequate control. Evidence will typically involve
  - evidence of traceability for the values of important control parameters prescribed by the method
  - evidence that uncertainties in these parameters are sufficiently small for the purpose
- It is important for laboratories to check at least the most critical false response rate for a qualitative test.
- It is reasonable to expect laboratories to be following published codes of best practice in qualitative testing where they are available.
- Quantitative (i.e. numerical) reports of uncertainties in qualitative test results should not generally be expected.



\*Eurachem position paper:  
Accreditation and Quality Assurance 5, 346–348(2000)